



## Expansion of Gamma Delta T Cells – A Short Review on Bisphosphonate and K562-Based Methods

Wee Kiat Tan<sup>1</sup>, Johan CK Tay<sup>2</sup>, Jieming Zeng<sup>3</sup>, Min Zheng<sup>4</sup>, Shu Wang<sup>2,3\*</sup>

<sup>1</sup>Tessa Therapeutics, Pte Ltd., Singapore 239351

<sup>2</sup>Department of Biological Sciences, National University of Singapore, Singapore 117543

<sup>3</sup>Institute of Bioengineering and Nanotechnology, Singapore 138669

<sup>4</sup>Department of Dermatology, Second Affiliated Hospital, Zhejiang University, School of Medicine, China 310009

### Article Info

#### Article Notes

Received: March 22, 2018

Accepted: May 25, 2018

#### \*Correspondence:

Dr. Shu Wang, Department of Biological Sciences National University of Singapore 14 Science Drive 4, Singapore 117543;  
Telephone: 65-6874 7712;  
Fax: 65-6779 2486;  
E-mail: dbsws@nus.edu.sg

© 2018 Wang S. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License.

### Gamma Delta T cells As the Next Generation of Effector Cells for Immunotherapy

Gamma delta ( $\gamma\delta$ )-T cells are thymus-derived lymphocytes that differ from  $\alpha\beta$ -T cells in their anatomical distributions and mechanisms of activation and function. They could be found as resident lymphocytes in human tissues or as circulating lymphocytes in peripheral blood<sup>1</sup> (Vantourout & Hayday, 2013). Unlike  $\alpha\beta$ -T cells that function exclusively in adaptive immunity,  $\gamma\delta$ -T cells are innate-like immune cells that recognize malignant cells through their repertoire of activating receptors in a MHC-independent manner, which is similar to natural killer cells<sup>2</sup> (Welsh et al., 1997).

Growing interest in the use of  $\gamma\delta$ -T cells as an alternative form of immunotherapy has been fueled partially by the MHC-restriction mechanism that limits  $\alpha\beta$ -T cell-based cancer immunotherapy to an autologous setting.  $\gamma\delta$ -T cells recognize antigens independently of MHC molecules and this mode of action avoids GvHD when used in an allogeneic setting<sup>3,4</sup> (Deniger et al., 2014; Xiao et al., 2018). Owing to their repertoire of activating receptors,  $\gamma\delta$ -T cells have been reported to kill a wide variety of cancer cell lines *in vitro* such as leukemia, melanoma, lymphomas and other carcinomas<sup>5</sup> (Yoshikawa et al., 2014). Drawing inspiration from the growing field of chimeric antigen receptors (CAR), mRNA CAR-engrafted  $\alpha\beta$ -T and  $\gamma\delta$ -T cells have been investigated in parallel and were found to exhibit comparable cytolytic activity against melanoma cells. However,  $\gamma\delta$ -T cells produced less proinflammatory cytokines as compared with  $\alpha\beta$ -T which make patients infused with  $\gamma\delta$ -T cells less susceptible to cytokine release syndrome<sup>6</sup> (Harrer et al., 2017). Given the above qualities,  $\gamma\delta$  T cells may be produced in large quantity as an off-the-shelf agent for cancer immunotherapy.

$\gamma\delta$ -T cells consist of 2 major subtypes, namely V $\delta$ 1 TCR and V $\delta$ 2 TCR subtype. Both subtypes could be found in the peripheral blood but the V $\delta$ 1 subtype is predominantly found in the epithelial tissues. The V $\delta$ 1 subtype reacts strongly to stress-associated factors commonly presented on cancer cells and virus-infected cells and act to destroy these cells, hence forming the first line of defense in the epithelial tissues<sup>7</sup> (Ebert, Meuter, & Moser, 2006). (Vantourout & Hayday, 2013) <sup>1</sup>Furthermore, V $\delta$ 1  $\gamma\delta$ -T cells have been found to directly kill Gram-positive bacteria found in the gut and promote wound healing through enhancing keratinocyte proliferation on the skin<sup>8</sup> (Nielsen, Witherden, & Havran, 2017).

In the peripheral blood,  $\gamma\delta$ -T cells only compose 0.5 to 5% of peripheral blood mononuclear cells (PBMCs) whereas  $\alpha\beta$ -T cells make up approximately 50%. Among these circulating  $\gamma\delta$ -T cells, V $\gamma$ 9V $\delta$ 2 T cells are the majority subset that has been well-studied for their anti-tumour activity<sup>9</sup> (Davodeau et al., 1993) but only account for less than 10% of all T cells<sup>10</sup> (Dopfer et al., 2014). The infrequent nature of  $\gamma\delta$ -T cells had therefore impeded their clinical application. However, the fortuitous finding in the 1990s that bisphosphonates could activate the *in vivo* proliferation of V $\gamma$ 9V $\delta$ 2 T cells during treatment of bone resorption pioneered the search for more expansion methods. On this basis, derivative methods such as cytokine-based and K562 feeder cell-based protocols have been developed to achieve clinically relevant numbers for cancer immunotherapy. In this review, we survey the current landscape of  $\gamma\delta$ -T cell culturing methods that address the need in V $\gamma$ 9V $\delta$ 2-based cancer immunotherapy.

### Bisphosphonate-Based Expansion of V $\gamma$ 9V $\delta$ 2 T Cells

Cytokine-based approaches for expansion of  $\gamma\delta$ -T cells have thus far revolved around the combinatorial use of both IL-2 and Zoledronate (ZOL) or the synthetic phosphoantigen bromohydrin pyrophosphate (BrHPP). ZOL and BrHPP share structural homology with non-peptide  $\gamma\delta$ -T cell ligands such as the isopentenyl pyrophosphate (IPP) intermediate found in the mevalonate pathway. The farnesyl pyrophosphate synthase (FPPS) enzyme in the mevalonate pathway converts IPP to farnesyl pyrophosphate, and the various metabolites generated through this pathway have been implicated in tumour growth and progression. Nitrogen-containing bisphosphonates (N-BPs) such as ZOL inhibit the FPPS enzyme through competitive inhibition<sup>11</sup> (Kavanagh et al., 2006) and though unclear, synthetic analogues such as BrHPP could be functioning in a similar fashion.

Addition of ZOL or BrHPP to PBMC culture blocks FPPS activity and causes an accumulation of IPP in monocytes<sup>12</sup> (Roelofs et al., 2009) and MHC Class II+ cells<sup>13</sup> (Soriano-Sarabia et al., 2012), which collectively activates V $\gamma$ 9V $\delta$ 2 T cells through cell-to-cell interactions. Although V $\gamma$ 9V $\delta$ 2 T cells can function as antigen-presenting cells (APCs) to CD8+ cytotoxic T lymphocytes (CTLs)<sup>5</sup> (Yoshikawa et al., 2014), it is unclear if they cross-present IPP for mutual activation. In earlier publications, V $\gamma$ 9V $\delta$ 2 T cells were purified from PBMCs before culturing with FPPS inhibitors and IL-2. As described by Salot et al., labelled V $\gamma$ 9V $\delta$ 2 T cells were sorted from PBMCs with magnetic beads; the unlabeled cell fraction was irradiated and used for co-culturing with the sorted V $\gamma$ 9V $\delta$ 2 T cells<sup>14</sup> (Salot et al., 2009). Although gamma irradiation may reduce concomitant expansion of non- $\gamma\delta$  T cells, antigen presenting capabilities of professional APCs could also be impaired by such treatment<sup>15</sup> (Merrick et al., 2005) which

may explain the low expansion folds of V $\gamma$ 9V $\delta$ 2 T cells using this method.

Besides APCs, CD4 T cells and their secreted cytokines also contribute to the activation of V $\gamma$ 9V $\delta$ 2 T cells. In a *Mycobacterium tuberculosis* model, V $\gamma$ 9V $\delta$ 2 T cells were primed by cytokines secreted by CD4 T cells such as IFN $\gamma$  and IL-2<sup>16</sup> (Pechhold, Wesch, Schondelmaier, & Kabelitz, 1994). The removal of CD4+ and CD8+ T cells may, therefore, have contributed to lowered expansion fold of the V $\gamma$ 9V $\delta$ 2 T cells<sup>17</sup> (Dokouhaki et al., 2010) although this could be partially rescued by exogenous IL-2.

In these cytokine-based approaches, the length of culture was often kept between 10 – 14 days (Table 1). V $\gamma$ 9V $\delta$ 2 T cells harvested from day 12 to day 14 might be most optimal for therapy as cytotoxic activity peaked on day 12 and declined by day 14<sup>18</sup> (Kondo et al., 2008). Prolonged exposure of V $\gamma$ 9V $\delta$ 2 T cells to phosphoantigens beyond 14 days caused activation-induced cell death (AICD)<sup>19</sup> (Ferrarini et al., 2008). Therefore, lengthening the culture period in pursuit of increased V $\gamma$ 9V $\delta$ 2 T cell expansion runs the risk of producing exhausted V $\gamma$ 9V $\delta$ 2 T cells with lower cytotoxic activity.

Although the highest rate of cytokine-based expansion reported by Kondo et al. stands at 13750 fold<sup>20</sup> (Kondo et al., 2011), all other methods reported a range of between 180 to 5000-fold (Table 1). Therefore, the infrequent nature of V $\gamma$ 9V $\delta$ 2 T cells necessitates a large starting number of PBMC to generate adequate effector V $\gamma$ 9V $\delta$ 2 T cells for ACT. Estimating at an average therapeutic dose of 2x10<sup>6</sup>  $\gamma\delta$ -T cells/kg bodyweight<sup>21</sup> (Wilhelm et al., 2014) and an average human body weight of 62kg<sup>22</sup> (Walpole et al., 2012), a typical ACT would require at least 1.2x10<sup>8</sup> cells per infusion. This means that leukapheresis, which is an invasive and strenuous procedure in order to obtain adequate starting PBMCs, becomes inevitable and hence complicating the process of V $\gamma$ 9V $\delta$ 2 T cell generation. In order to develop an off-the-shelf cancer immunotherapy, a larger expansion rate of the V $\gamma$ 9V $\delta$ 2 T cell is needed to streamline the process and lower the cost of production.

### Feeder Cell-Based V $\gamma$ 9V $\delta$ 2 T Cells Expansion Methods for Clinical Use

The various reports reviewed thus far have highlighted a need to conserve the non- $\gamma\delta$  T cell fraction during activation by Zol and IL-2, and a need to consider the duration of culture such that a clinically-sufficient number of V $\gamma$ 9V $\delta$ 2 T cell could be obtained for adoptive cell therapy.

To generate even larger numbers of effector cells, artificial antigen-presenting cells (aAPCs) have been explored as a feeder cell-based method to provide a more sustained source of activation and costimulation. K562 is an MHC-negative human chronic erythroleukemic cell line

that has been genetically modified and adapted for cell-based expansion of polyclonal CTLs<sup>23</sup> (Maus et al., 2002) and natural killer cells<sup>24</sup> (Imai, Iwamoto, & Campana, 2005). K562 was initially chosen as a cellular scaffold for genetic modification because the lack of MHC class I and class II expression prevents allogeneic responses from donor T lymphocytes<sup>23</sup> (Maus et al., 2002).

Deniger and colleagues were the first group to adopt this aAPC technology for the expansion of polyclonal  $\gamma\delta$ -T cells<sup>3</sup> (Deniger et al., 2014) (Table 2). After the use of microbeads to remove NK cells and positively-select for polyclonal  $\gamma\delta$ -T cells, the authors used K562 feeder cells, modified to express CD19, CD64, CD86, CD137L and membrane-bound IL15, to expand and maintain the polyclonal  $\gamma\delta$ -T populations. These K562 cells were able to induce an expansion of  $4900 \pm 1700$  folds (Table 2), which is a clear improvement from figures obtained from cytokine-based methods (Table 1). Unlike two later papers that focused on the V $\gamma$ 9V $\delta$ 2 subset<sup>25,4</sup> (Cho et al., 2016; Xiao et al., 2018), Deniger used these feeder cells to expand and study the polyclonal population and also the isolated V $\delta$ 1 and V $\delta$ 2 subsets, and interestingly found that the previously less-studied V $\delta$ 1 subset was capable of elucidating anti-tumour

effects as well. In addition, the use of K562 cells as aAPC seemed to negate the AICD effects of prolonged exposure to antigens. Over a 35-day co-culture period, Deniger reported up to  $10^7$ -fold increase in umbilical cord blood (UCB)-derived  $\gamma\delta$ -T cells<sup>3</sup> (Deniger et al., 2014). The authors opined that this application is potentially useful as the younger UCB-derived  $\gamma\delta$ -T cells contain a more diverse repertoire of TCR $\gamma\delta$  and would support a longer-term engraftment in allogeneic recipient patients.

Also working on V $\gamma$ 9V $\delta$ 2 T cells, Cho and colleagues used a different K562 cell line that was modified to express CD32, CD83 and CD137L instead to expand V $\gamma$ 9V $\delta$ 2 T cells purified from PBMCs by FACS sorting. Unlike the expansion figures achieved by Deniger et al., Cho and colleagues were only able to achieve an approximately 12-fold expansion by day 14 and a  $10^6$ -fold by day 100<sup>25</sup> (Cho et al., 2016). In terms of clinical applications, this adds significantly to the manufacturing costs and duration.

Concerns with the use of K562 feeder cells for immune cell expansion lie primarily with the potential presence of residual K562 cells after co-culture and post-infusion into patients. To address this, K562 cells are gamma-irradiated to arrest their cell growth and to prevent unintended in

Cytokines and Reagents Used	Duration (Days)	$\gamma\delta$ -T Expansion Fold	Nature of Sorting Procedure Performed	Reference
IFN $\gamma$ , IL-12, anti-CD2 mAb, anti-CD3 mAb (OKT3), IL-2	14	Not determined; 70-fold change for total cell count	After initial expansion	(Liu, Guo, Gehrs, Nan, & Lopez, 2005) <sup>34</sup>
BrHPP, IL-2, OKT3	14	1585 $\pm$ 1493	No	(Salot et al., 2007) <sup>35</sup>
ZOL, IL-2	14	4798 $\pm$ 654	No	(Kondo et al., 2008) <sup>18</sup>
BrHPP, IL-2, OKT3	10	180	PBMCs were sorted with MACS "TCR $\gamma/\delta$ + T cell isolation kit" before coculture with irradiated remainder of PBMCs	(Salot et al., 2009) <sup>14</sup>
IL-2, IL-4, OKT3	10 to 14	376 $\pm$ 185	CD4 and CD8 T cell depletion with RosetteSep depletion kit before culture	(Dokouhaki et al., 2010) <sup>17</sup>
ZOL, IL-2, IL-18	14	1000-5000	N.D	(Li et al., 2010) <sup>36</sup>
ZOL, IL-2	10 to 14	13750	No	(Kondo et al., 2011) <sup>20</sup>
ZOL, IL-2, IL-15	14	1000	No	(Van Acker et al., 2016) <sup>37</sup>

**Table 1:** Comparison of bisphosphonate and cytokine based methods for  $\gamma\delta$ -T cell expansion

Cytokines and Reagents Used	$\gamma\delta$ -T Expansion Fold	Co-Culture Ratio of T Cell to Feeder Cell	Genetic Modification of K562 Feeder Cells	Frequency of Stimulation with Feeder Cells	Nature of Sorting Performed	Reference
IL-2, IL-21	4900 $\pm$ 1700	1 to 2	CD19, CD64, CD86, CD137L, membrane-bound IL-15	every 7 days	CD56 microbeads for removal of NK cells; TCR $\gamma/\delta$ + T cell isolation kit for positive selection of $\gamma\delta$ -T cells	(Deniger et al., 2014) <sup>3</sup>
IL-2, $\alpha$ CD3, $\alpha$ CD28	12-fold at day 14, 1,000,000-fold by day 100	2 to 1	CD32, CD80, CD137L	every 7 days	FACS-based positive selection of V $\gamma$ 9V $\delta$ 2 T cells prior to co-culture with aAPCs	(Cho et al., 2016) <sup>25</sup>
ZOL, IL-2, OKT3	537340 $\pm$ 607389	1 to 100	CD64, CD86, CD137L	once on day 7	Magnetic bead-based removal of $\alpha\beta$ -T cells from day 7 culture before stage 2 culture with K562 aAPCs	(Xiao et al., 2018) <sup>4</sup>

**Table 2:** Comparison of K562-based  $\gamma\delta$ -T cell expansion methods

vivo proliferation after ACT. Various studies, including ours, have reported very low or undetectable levels of gamma-irradiated K562 feeder cells after 7 days of co-culture<sup>26,4</sup> (Lapteva et al., 2012; Xiao et al., 2018). FDA regulations require that there should be less than 0.1% of detectable gamma-irradiated feeder cells in the final product<sup>26</sup> (Lapteva et al., 2012). As such, immune effector cells expanded by K562 feeder cells have been approved for clinical trials by the FDA (NKEXP and NKCD19; clinicaltrials.gov numbers NCT00640796 and NCT00995137).

### Combinatorial Usage of ZOL, OKT3 and K562 Feeder Cells in an FDA-Registered G-Rex Device

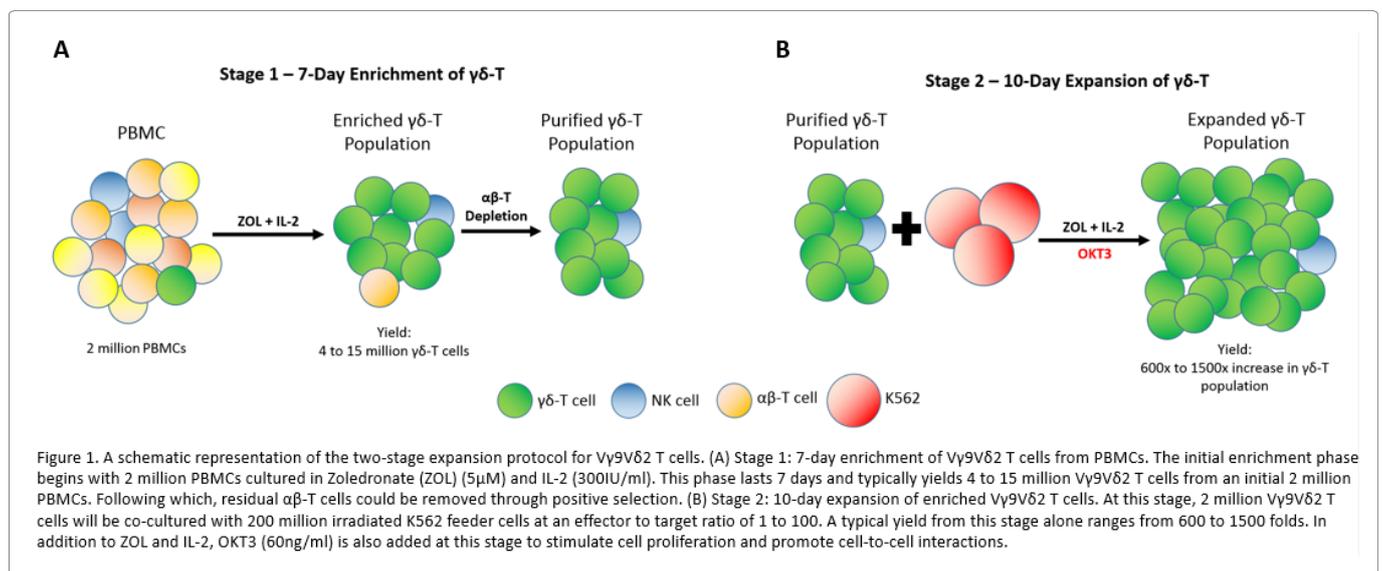
In our work, we had chosen to focus on the V $\gamma$ 9V $\delta$ 2 T cell population because it is the subset that has been most extensively studied for its safety and efficacy<sup>27,28,29</sup> (Fisher, Heuijerjans, Yan, Gustafsson, & Anderson, 2014; Fournie et al., 2013; Kobayashi & Tanaka, 2015). This sets our work apart from that performed by Deniger although their expansion protocol for a polyclonal  $\gamma\delta$ -T cell population remains impressive. The key factor that differentiates our work from Cho et al. then is the initial stage of enriching V $\gamma$ 9V $\delta$ 2 T cell from the general PBMC population by ZOL and IL-2 as compared to cell sorting. As explained in the previous section, the use of ZOL to enrich V $\gamma$ 9V $\delta$ 2 T cell has been well-validated and requires the presence of accessory cells such as monocytes and other MHC class II-positive cells. The use of K562 feeder cells, as demonstrated by Deniger et al., does not seem to favour any particular  $\gamma\delta$ -T subset and merely serves to numerically expand the population. The two-stage protocol developed in our paper, therefore, leverages on the ability of ZOL to enrich V $\gamma$ 9V $\delta$ 2 T cells, and the ability of K562 feeder cells to numerically expand the enriched population.

In our two-stage protocol (Figure 1), a robust and selective activation of V $\gamma$ 9V $\delta$ 2 T cells was achieved in the

first phase through the use of ZOL prior to sorting<sup>4</sup> (Xiao et al., 2018). A well-timed magnetic sorting at day 7 after activation would remove NK and  $\alpha\beta$  T cell contaminants but maintain the activation status of V $\gamma$ 9V $\delta$ 2 T cells. This is evident in the potent fold expansion of V $\gamma$ 9V $\delta$ 2 T cells of  $537340 \pm 607389$  in 17 days. This is the fastest and highest V $\gamma$ 9V $\delta$ 2 T cell expansion figure we have observed to date. Although there may be concern arising from the high ratio of V $\gamma$ 9V $\delta$ 2 T to K562 used at 1:100, we could prove minimal residual feeder cells at day 17, and that the overall usage of K562 feeder cells was limited to once for the whole duration.

The fact that K562 cells could support a longer-term *in vitro* culture, as compared to the typical 10-14 day period in cytokine-based methods, was also demonstrated in our own study<sup>4</sup> (Xiao et al., 2018). We had reported that the 10-day co-culture period with K562 brought about a decrease in classic T cell exhaustion markers such as CTLA-4, LAG3, TIM3 and BTLA4. Day-17 V $\gamma$ 9V $\delta$ 2 T cells were activated and highly cytotoxic against ZOL-treated cancer cells. Cytotoxicity assays also did not reveal any difference between day 7 ZOL/IL-2-expanded V $\gamma$ 9V $\delta$ 2 T cells and day 17 K562-expanded V $\gamma$ 9V $\delta$ 2 T cells (unpublished data). Although there was no observable *in vitro* difference in cytotoxicity, the down-regulated expression of immune checkpoint inhibitors could possibly nullify the strongly immunosuppressive tumour microenvironment and maintain *in vivo* anti-tumour activity.

The huge numerical expansion is explained by the usage of anti-CD3 antibody OKT3 that targets the CD3 epsilon chain expressed on  $\gamma\delta$ -T cells. Importantly, we built on the interesting observation by Dopfer et al. that OKT3 could promote significant cytokine secretion and proliferation by V $\gamma$ 9V $\delta$ 2 T cells<sup>10</sup> (Dopfer et al., 2014). Our K562 feeder cell line is engineered to express CD64, which



is a high-affinity human IgG FcγRI receptor. The interaction between OKT3 and CD64 thus promotes cell-to-cell contact between K562 and Vγ9Vδ2 T cells, which in turn enables better interactions between CD86 and CD137L on expressed on our K562 feeder cell line and the CD28 and CD137 costimulatory molecules expressed on Vγ9Vδ2 T cells respectively.

To set the stage for future clinical trials and applications with these K562-expanded Vγ9Vδ2 T cells, we trialed the use of the G-Rex (Wilson Wolf, US) system in the second stage of our expansion protocol. G-Rex is designed to support cell cultures with low cell seeding density at  $0.125 \times 10^6 / \text{cm}^2$ <sup>30</sup> (Bajgain et al., 2014). Its key feature is the gas-permeable silicone membrane at the bottom that facilitates efficient exchange of O<sub>2</sub> and CO<sub>2</sub> which enables more media to be added per unit of surface area. By doing so, the frequency of media exchange is reduced, and the increased nutrient support also enhances cell survival<sup>31</sup> (Vera et al., 2010). Its compact design and lower requirement for manpower also enable scalability in large-scale cell manufacturing which is an important consideration for commercialization. In our work, we seeded  $1 \times 10^6$  day 7 ZOL-activated Vγ9Vδ2 T cells with  $1 \times 10^8$  irradiated-K562 feeder cells at a coculture ratio of 1:100<sup>4</sup> (Xiao et al., 2018). With this FDA-registered class I medical device, we could achieve an expansion of between 1,000 to 15,000 folds in just 10 days. The use of G-Rex thus enables us to achieve a clinically-relevant number of cells needed for effective therapeutic intervention.

### The Role of Vγ9Vδ2 T Cells in Th1 Cytokine Secretion and Chimeric Antigen Receptor

Recently, there has been emerging concern about the pathogenic nature of certain γδ-T cells. Indeed, it has been noted that polyclonal γδ-T cells expanded by K562 aAPC secrete Th17 cytokines including IL-17 when activated by a leukocyte activation cocktail<sup>3</sup> (Deniger et al., 2014). Yet, in the same experiment, it was shown that there was a significant secretion of interferon-gamma (IFN-γ), which is a good surrogate marker for cytotoxicity. Additional experiments had shown that the Vδ2 subset was the main source of IFN-γ, indicating that Vγ9Vδ2 T cells might be the majority population of the polyclonal γδ-T cells. Another review pointed out that γδ-T cells may present Th2 or Th17 profile based on the cytokine milieu within the microenvironment. Yet, these findings are largely based on results from *in vitro* experiments and not pre-clinical models<sup>32</sup> (Xiang & Tu, 2017). This remains a concern and more preclinical data should be collected to understand the mechanism behind γδ-T cells adopting a Th17 profile in certain tumour microenvironment. On our part, we have mitigated this risk by focusing on the Vγ9Vδ2 subset, which is known to express a Th1 signature with the majority producing proinflammatory IFN-γ and a minority (<5%) secreting IL-17<sup>33</sup> (Patil, Bhat, Dar, & Chiplunkar, 2015).

The combined use of ZOL and Vγ9Vδ2 T cells presents clear therapeutic effect in clinical trials with patients achieving partial remission in advanced breast and prostate cancer and complete remission of RCC patient with lung metastasis. However, certain cancer cells are resistant to ZOL treatment and do not respond well to Vγ9Vδ2 T cells. To enhance the anti-tumour activity of these K562-expanded Vγ9Vδ2 T cells, we have shown in our work that CAR-engrafted Vγ9Vδ2 T cells function efficiently against EpCAM-positive cancer cells. Given their propensity to home in to epithelial tissues and the antigen-specificity that could be afforded by CAR technology, γδ-T cells have huge potential for clinical applications against various solid tumours.

### Conclusion and Future Perspective

A significant milestone in gene therapy and adoptive cell therapy was achieved with the recent FDA approval of CAR-T cells for use in relapsed B-cell malignancies. However, the dependence on autologous αβ-T cells to drive this anti-tumour effort has placed a huge constraint on logistics and supply chain management, from individualized sample collection from patient, to the *ex vivo* expansion, genetic modification and quality control. This has considerably driven up costs and time needed for effective therapeutic interventions. In this aspect, Vγ9Vδ2 T cells represent a hugely viable option to replace conventional αβ-T cells as an off-the-shelf immunotherapeutic agent. The intrinsic anti-tumour activity of γδ cells, coupled with their MHC-independent modus operandi, makes them ideal allogenic off-the-shelf effector cells for cancer immunotherapy. The bottleneck had been the large-scale expansion of these scarce cells, and this has been overcome to various extents by the various methods reported. We believe that future clinical trials involving Vγ9Vδ2 T cells would explore the combinatorial use with other therapeutic agents, such as CAR and immune checkpoint inhibitors. With our recent publication<sup>4</sup> (Xiao et al., 2018), we have provided an economical method to achieve large-scale expansion of these Vγ9Vδ2 T cells and have also validated their combinatorial use with CAR technology. Our use of CD64, CD86 and CD137L-expressing K562 feeder cells has also mitigated the risks of having exhausted Vγ9Vδ2 T cells from an extended period of *in vitro* culture. Future efforts in our laboratory and others should strive to constantly improve the quality of the expanded Vγ9Vδ2 T cells while reducing the cost of therapy, thereby allowing the therapy to reach the masses efficiently.

### References

1. Vantourout P, Hayday A. Six-of-the-best: unique contributions of gammadelta T cells to immunology. Nat Rev Immunol. 2013; 13(2): 88-100. doi:10.1038/nri3384
2. Welsh RM, Lin MY, Lohman BL, et al. Alpha beta and gamma delta T-cell networks and their roles in natural resistance to viral infections. Immunol Rev. 1997; 159: 79-93.

3. Deniger DC, Maiti SN, Mi T, et al. Activating and propagating polyclonal gamma delta T cells with broad specificity for malignancies. *Clin Cancer Res.* 2014; 20(22): 5708-5719. doi:10.1158/1078-0432.CCR-13-3451
4. Xiao L, Chen C, Li Z, et al. Large-scale expansion of Vgamma9Vdelta2 T cells with engineered K562 feeder cells in G-Rex vessels and their use as chimeric antigen receptor-modified effector cells. *Cytotherapy.* 2018; 20(3): 420-435. doi:10.1016/j.jcyt.2017.12.014
5. Yoshikawa T, Takahara M, Tomiyama M, et al. Large-scale expansion of gammadelta T cells and peptide-specific cytotoxic T cells using zoledronate for adoptive immunotherapy. *Int J Oncol.* 2014; 45(5): 1847-1856. doi:10.3892/ijo.2014.2634
6. Harrer DC, Simon B, Fujii SI, et al. RNA-transfection of gamma/delta T cells with a chimeric antigen receptor or an alpha/beta T-cell receptor: a safer alternative to genetically engineered alpha/beta T cells for the immunotherapy of melanoma. *BMC Cancer.* 2017; 17(1): 551. doi:10.1186/s12885-017-3539-3
7. Ebert LM, Meuter S, Moser B. Homing and Function of Human Skin T Cells and NK Cells: Relevance for Tumor Surveillance. *The Journal of Immunology.* 2006; 176(7): 4331-4336. doi:10.4049/jimmunol.176.7.4331
8. Nielsen MM, Witherden DA, Havran WL. gammadelta T cells in homeostasis and host defence of epithelial barrier tissues. *Nat Rev Immunol.* 2017; 17(12): 733-745. doi:10.1038/nri.2017.101
9. Davodeau F, Peyrat MA, Hallet MM, et al. Peripheral selection of antigen receptor junctional features in a major human gamma delta subset. *Eur J Immunol.* 1993; 23(4): 804-808. doi:10.1002/eji.1830230405
10. Dopfer EP, Hartl FA, Oberg HH, et al. The CD3 conformational change in the gammadelta T cell receptor is not triggered by antigens but can be enforced to enhance tumor killing. *Cell Rep.* 2014; 7(5): 1704-1715. doi:10.1016/j.celrep.2014.04.049
11. Kavanagh KL, Guo K, Dunford JE, et al. The molecular mechanism of nitrogen-containing bisphosphonates as antiosteoporosis drugs. *Proc Natl Acad Sci U S A.* 2006; 103(20): 7829-7834. doi:10.1073/pnas.0601643103
12. Roelofs AJ, Jauhainen M, Monkkonen H, et al. Peripheral blood monocytes are responsible for gammadelta T cell activation induced by zoledronic acid through accumulation of IPP/DMAPP. *Br J Haematol.* 2009; 144(2): 245-250. doi:10.1111/j.1365-2141.2008.07435.x
13. Soriano-Sarabia N, Sandvold H, Jomaa H, et al. Primary MHC-class II(+) cells are necessary to promote resting Vdelta2 cell expansion in response to (E)-4-hydroxy-3-methyl-but-2-enyl-pyrophosphate and isopentenyl pyrophosphate. *J Immunol.* 2012; 189(11): 5212-5222. doi:10.4049/jimmunol.1200093
14. Salot S, Bercegeay S, Dreno B, et al. Large scale expansion of Vgamma9Vdelta2 T lymphocytes from human peripheral blood mononuclear cells after a positive selection using MACS "TCR gamma/delta+ T cell isolation kit". *J Immunol Methods.* 2009; 347(1-2): 12-18. doi:10.1016/j.jim.2009.05.006
15. Merrick A, Errington F, Milward K, et al. Immunosuppressive effects of radiation on human dendritic cells: reduced IL-12 production on activation and impairment of naive T-cell priming. *Br J Cancer.* 2005; 92(8): 1450-1458. doi:10.1038/sj.bjc.6602518
16. Pechhold K, Wesch D, Schondelmaier S, et al. Primary activation of V gamma 9-expressing gamma delta T cells by Mycobacterium tuberculosis. Requirement for Th1-type CD4 T cell help and inhibition by IL-10. *J Immunol.* 1994; 152(10): 4984-4992.
17. Dokouhaki P, Han M, Joe B, et al. Adoptive immunotherapy of cancer using ex vivo expanded human gamma delta T cells: A new approach. *Cancer Letters.* 2010; 297(1): 126-136. doi:10.1016/j.canlet.2010.05.005
18. Kondo M, Sakuta K, Noguchi A, et al. Zoledronate facilitates large-scale ex vivo expansion of functional gammadelta T cells from cancer patients for use in adoptive immunotherapy. *Cytotherapy.* 2008; 10(8): 842-856. doi:10.1080/14653240802419328
19. Ferrarini M, Delfanti F, Gianolini M, et al. NF-kappa B modulates sensitivity to apoptosis, proinflammatory and migratory potential in short- versus long-term cultured human gamma delta lymphocytes. *J Immunol.* 2008; 181(9): 5857-5864.
20. Kondo M, Izumi T, Fujieda N, et al. Expansion of human peripheral blood gammadelta T cells using zoledronate. *J Vis Exp.* 2011; (55). doi:10.3791/3182
21. Wilhelm M, Smetak M, Schaefer-Eckart K, et al. Successful adoptive transfer and in vivo expansion of haploidentical  $\gamma\delta$  T cells. *Journal of Translational Medicine.* 2014; 12(1): 45. doi:10.1186/1479-5876-12-45
22. Walpole SC, Prieto-Merino D, Edwards P, et al. The weight of nations: an estimation of adult human biomass. *BMC Public Health.* 2012; 12(1): 439. doi:10.1186/1471-2458-12-439
23. Maus MV, Thomas AK, Leonard DG, et al. Ex vivo expansion of polyclonal and antigen-specific cytotoxic T lymphocytes by artificial APCs expressing ligands for the T-cell receptor, CD28 and 4-1BB. *Nat Biotechnol.* 2002; 20(2): 143-148. doi:10.1038/nbt0202-143
24. Imai C, Iwamoto S, Campana D. Genetic modification of primary natural killer cells overcomes inhibitory signals and induces specific killing of leukemic cells. *Blood.* 2005; 106(1): 376-383. doi:10.1182/blood-2004-12-4797
25. Cho HW, Kim SY, Sohn DH, et al. Triple costimulation via CD80, 4-1BB, and CD83 ligand elicits the long-term growth of Vgamma9Vdelta2 T cells in low levels of IL-2. *J Leukoc Biol.* 2016; 99(4): 521-529. doi:10.1189/jlb.1HI0814-409RR
26. Lapteva N, Durett AG, Sun J, et al. Large-scale ex vivo expansion and characterization of natural killer cells for clinical applications. *Cytotherapy.* 2012; 14(9): 1131-1143. doi:10.3109/14653249.2012.700767
27. Fisher JP, Heuveljans J, Yan M, et al. gammadelta T cells for cancer immunotherapy: A systematic review of clinical trials. *Oncoimmunology.* 2014; 3(1): e27572. doi:10.4161/onci.27572
28. Fournie JJ, Sicard H, Poupot M, et al. What lessons can be learned from gammadelta T cell-based cancer immunotherapy trials. *Cell Mol Immunol.* 2013; 10(1): 35-41. doi:10.1038/cmi.2012.39
29. Kobayashi H, Tanaka Y. gammadelta T Cell Immunotherapy-A Review. *Pharmaceuticals (Basel).* 2015; 8(1): 40-61. doi:10.3390/ph8010040
30. Bajgain P, Mucharla R, Wilson J, et al. Optimizing the production of suspension cells using the G-Rex "M" series. *Mol Ther Methods Clin Dev.* 2014; 1: 14015. doi:10.1038/mtm.2014.15
31. Vera JF, Brenner LJ, Gerdemann U, et al. Accelerated production of antigen-specific T cells for preclinical and clinical applications using gas-permeable rapid expansion cultureware (G-Rex). *J Immunother.* 2010; 33(3): 305-315. doi:10.1097/CJI.0b013e3181c0c3cb
32. Xiang Z, Tu W. Dual Face of V $\gamma$ 9V $\delta$ 2 T Cells in Tumor Immunology: Anti- versus Pro-Tumoral Activities. *Frontiers in Immunology.* 2017; 8: 1041. doi:10.3389/fimmu.2017.01041
33. Patil RS, Bhat SA, Dar AA, et al. The Jekyll and Hyde story of IL17-Producing gammadelta T Cells. *Front Immunol.* 2015; 6: 37. doi:10.3389/fimmu.2015.00037
34. Liu Z, Guo BL, Gehrs BC, et al. Ex vivo expanded human Vgamma9Vdelta2+ gammadelta-T cells mediate innate antitumor activity against human prostate cancer cells in vitro. *J Urol.* 2005; 173(5): 1552-1556.
35. Salot S, Laplace C, Saiagh S, et al. Large scale expansion of gamma 9 delta 2 T lymphocytes: Innacell gamma delta cell therapy product. *J Immunol Methods.* 2007; 326(1-2): 63-75. doi:10.1016/j.jim.2007.07.010

36. Li W, Kubo S, Okuda A, et al. Effect of IL-18 on expansion of gammadelta T cells stimulated by zoledronate and IL-2. *J Immunother.* 2010; 33(3): 287-296. doi:10.1097/CJI.0b013e3181c80ffa
37. Van Acker HH, Anguille S, Willems Y, et al. Interleukin-15 enhances the proliferation, stimulatory phenotype, and antitumor effector functions of human gamma delta T cells. *J Hematol Oncol.* 2016; 9(1): 101. doi:10.1186/s13045-016-0329-3